Magnetic resonance spectroscopy of paragangliomas: new insights into in vivo metabolomics

To cite this version:

HAL Id: hal-01414336
https://hal-amu.archives-ouvertes.fr/hal-01414336
Submitted on 9 Dec 2019

HAL is a multi-disciplinary open access archive for the deposit and dissemination of scientific research documents, whether they are published or not. The documents may come from teaching and research institutions in France or abroad, or from public or private research centers. L’archive ouverte pluridisciplinaire HAL, est destinée au dépôt et à la diffusion de documents scientifiques de niveau recherche, publiés ou non, émanant des établissements d’enseignement et de recherche français ou étrangers, des laboratoires publics ou privés.

Distributed under a Creative Commons Attribution 4.0 International License
Magnetic resonance spectroscopy of paragangliomas: new insights into in vivo metabolomics

Arthur Varoquaux1, Yann le Fur2, Alessio Imperiale3,4, Antony Reyre1, Marion Montava5, Nicolas Fakhry6, Izzie-Jacques Namer3,4, Guy Moulin1, Karel Pacak7, Maxime Guye1,2 and David Taieb8,9,10

1Department of Medical Imaging, La Timone University Hospital, Aix-Marseille University, Marseille, France
2CNRS, CRMBM UMR 7339, Aix-Marseille University, 13385 Marseille, France
3Department of Biophysics and Nuclear Medicine, University Hospitals of Strasbourg, Strasbourg, France
4Cube, UMR 7357, Faculty of Medicine, University of Strasbourg/CNRS and FMTS, Strasbourg, France
5Department of Otorhinolaryngology–Head and Neck Surgery, North Hospital, Aix-Marseille University, Marseille, France
6Department of Otorhinolaryngology–Head and Neck Surgery, La Timone University Hospital, Aix-Marseille University, Marseille, France
7Program in Reproductive and Adult Endocrinology, Eunice Kennedy Shriver National Institute of Child Health and Human Development (NICHD), National Institutes of Health, Bethesda, Maryland 20892, USA
8Department of Nuclear Medicine, La Timone University Hospital, Aix-Marseille University, Marseille, France
9Biophysics and Nuclear Medicine, European Center for Research in Medical Imaging, La Timone University Hospital, Aix-Marseille University, 264, Rue Saint-Pierre, 13385 Marseille, France
10INSERM, UMR1068, Institut Paoli-Calmettes, Marseille Cancerology Research Center, Marseille, France

Correspondence should be addressed to D Taieb or A Varoquaux
Emails david.taieb@ap-hm.fr or DamienArthur.VAROQUAUX@ap-hm.fr

Abstract

Paragangliomas (PGLs) can be associated with mutations in genes of the tricarboxylic acid (TCA) cycle. Succinate dehydrogenase (SDHx) mutations are the prime examples of genetically determined TCA cycle defects with accumulation of succinate. Succinate, which acts as an oncometabolite, can be detected by ex vivo metabolomics approaches. The aim of this study was to evaluate the potential role of proton magnetic resonance (MR) spectroscopy (1H-MRS) for identifying SDHx-related PGLs in vivo and noninvasively. Eight patients were prospectively evaluated with single voxel 1H-MRS. MR spectra from eight tumors (four SDHx-related PGLs, two sporadic PGLs, one cervical schwannoma, and one cervical neurofibroma) were acquired and interpreted qualitatively. Compared to other tumors, a succinate resonance peak was detected only in SDHx-related tumor patients. Spectra quality was considered good in three cases, medium in two cases, poor in two cases, and uninterpretable in the latter case. Smaller lesions had lower spectra quality compared to larger lesions. Jugular PGLs also exhibited a poorer spectra quality compared to other locations. 1H-MRS has always been challenging in terms of its technical requisites. This is even more true for the evaluation of head and neck tumors. However, 1H-MRS might be added to the classical MR sequences for metabolomic characterization of PGLs. In vivo detection of succinate might guide genetic testing, characterize SDHx variants of unknown significance (in the absence of available tumor sample), and even optimize a selection of appropriate therapies.
**Introduction**

Paragangliomas (PGLs) are slow-growing hypervascular tumors arising from neural crest cell derivatives throughout the body. PGLs are closely aligned with the distribution of the autonomic nervous system and preferentially arise in the adrenal medulla, along the thoracolumbar sympathetic plexus, in the retroperitoneum, or in paragangliomas that are mainly located in the head and neck (Taieb et al. 2014a).

Approximately 30–40% of PGLs carry a germ line mutation, which frequently occurs in one of the succinate dehydrogenase (SDH) subunit genes (collectively referred to as SDHx) (Baysal et al. 2002, Neumann et al. 2009, Piccini et al. 2012, Martucci & Pacak 2014).

The SDH complex (also named mitochondrial complex II) catalyzes the oxidation of succinate to fumarate in the tricarboxylic acid (TCA) cycle and the respiratory chain. Deleterious mutations in any of the SDH genes (after biallelic inactivation) invariably result in decreased SDH activity, with accumulation of succinate, which acts as an oncometabolite (Selak et al. 2005).

We, and others, have recently shown that ex vivo metabolomics studies are very reliable methods for classifying various pheochromocytomas (PHEOs)/PGLs according to their genetic background (Imperiale et al. 2013a, Richter et al. 2014, Rao et al. 2015). Assessment of succinate concentration and succinate:fumarate ratio can be clinically relevant for discriminating SDHx-related tumors from sporadic and other hereditary PHEOs/PGLs (Lendvai et al. 2014, Richter et al. 2014, Imperiale et al. 2015a). These studies nicely pointed toward the importance of metabolite profiling in the evaluation of these tumors.

In recent years, anatomic and functional imaging techniques have gained an increasing role in the characterization of PHEOs/PGLs (Taieb et al. 2013, 2014a). The use of magnetic resonance imaging (MRI) as a non-ionizing technique is rapidly growing in the evaluation of PGLs with clinical implementation of multiparametric sequences that provide relevant biological information (i.e., diffusion weighted imaging, dynamic contrast enhancement, and spectroscopy). MR spectroscopy (MRS) enables quantification of metabolites in tissues (Bruhn et al. 1989, King et al. 2010). MRS uses intrinsic MR properties of some atomic nuclei (i.e., $^1$Hydrogen, $^{31}$Phosphorous, $^{19}$Fluorine, and $^{13}$Carbon) placed in a radiofrequency range of magnetic fields (de Graaf 2008). Proton spectroscopy ($^1$H-MRS) is available in numerous magnetic field strengths (currently from 1.5 to 7 Tesla) and has been evaluated in the characterization of various brain (Fellah et al. 2013) and extracerebral tumors (King et al. 2005, Jansen et al. 2012, Abdel Razek & Poptani 2013).

Currently, there is a strong interest in i) assessing the genetic and metabolomics backgrounds of tumors based on their metabolomics profile using noninvasive techniques that would not require obtaining additional tumor samples (in some patients, especially those with metastatic PHEO/PGL, it is difficult to obtain because biopsy may be contraindicated); ii) decreasing radiation exposure of cancer patients to the repeated use of anatomical and functional modalities in assessing or monitoring therapeutic responses; iii) minimizing the cost to a patient as well as the health care system by using multiple imaging modalities; and iv) selecting appropriate treatment options that are expected in the near future to be largely based on the assessment of tumor metabolomics profiles because metabolites are now considered as ‘first-line’ combat soldiers in a cancer cell.

Thus, the aim of the present study was to evaluate the potential role of 3T proton MRS in identifying various SDHx-related PGLs and then to compare those results to other sporadic PGLs or non-PGL tumors.

**Materials and methods**

**Patients**

Eight consecutive patients with suspicion of either HNPGL or a neck nerve sheath tumor were evaluated by $^1$H-MRS in addition to conventional MR sequences. PGL patients were included in a large prospective clinical trial dedicated to positron emission tomography (PET) imaging studies (NCT02186678) and were therefore evaluated by $^{18}$F-FDOPA and $^{68}$Ga-DOTATATE PET/computed tomography (CT) (patient nos 1–6) using low-dose CT protocol. The remaining two patients gave their informed consent for use of their personal data for scientific purposes, in keeping with local institutional guidelines.

**MRI protocol**

MRS was performed on a 3T MR scanner (Magnetom Skyra, Siemens Healthcare, Erlangen, Germany) equipped with a 32-channel phased-array head coil. The signal of monooxyl MRS was collected following the unenhanced conventional MR sequences (for head and neck regions: $T_1$, $T_2$, and time of flight angiography (TOF)). The volume of interest (VOI) was carefully positioned by a radiologist with 10 years’ experience (A Varoquaux). The VOI was adapted to the size...
and geometry and centered within the bulk tumor region, excluding nearby bone structures. Six outer-volume lipid suppression bands were used to suppress lipid contamination, and pre-acquisition included shimming and water suppression. Spectra were acquired with a point-resolved spectroscopy sequence (PRESS; TE, 135 ms and repetition time (TR), 2000 ms) using the manufacturer’s automated shimming procedure. An H2O signal was acquired for quantification purposes at the same location. Extra scanning time, including shimming and acquisition (120 excitations), was 8 min.

Post processing

The MRS data were analyzed using a dedicated software described elsewhere (Le Fur et al. 2010). After Fourier transformation, the residual water signal was removed using HLSVD (de Beer et al. 1992). Spectra was fitted using HRQUEST (Ratiney et al. 2005) with a simulated database that incorporated seven metabolites selected according to the HRMAS spectrum (Imperiale et al. 2013a,b, 2015a): acetate, alanine, glutamate, glutathione, lactate, methionine, and succinate.

Quality of the spectra were classified as follows: i) good: thin resonances, good water suppression, and absence of lipid signal contamination; ii) medium: broad resonances but clearly distinguishable, acceptable water suppression, and low lipid contamination; iii) poor: resonances hardly distinguishable and/or bad water suppression and/or high lipid signal; and iv) uninterpretable spectrum. MRS spectra were interpreted by experts blinded to the SDH mutation status and pathological findings.

HRMAS MRS

HRMAS MRS was performed in three cases (carotid body PGL (CBP) from patient nos 3 and 4 and one abdominal extra-adrenal PGL from patient no. 1) from the analysis of a frozen intact tumor sample of about 15 mg. Spectra were acquired on a Bruker Avance III 500 Spectrometer (500.13 MHz) (Bruker BioSpin, Wissembourg, France). One-dimensional (1D) proton and two-dimensional (2D) heteronuclear (1H-13C) experiments were recorded. Selected metabolites were quantified according to our previous reports (Imperiale et al. 2013b).

Gold standard

Pathological analysis of the tumor was considered the gold standard for final diagnosis. When surgery was not indicated or already performed, lesions were characterized as PGL by tumor positivity on either 18F-FDOPA or 68Ga-DOTATATE in specific locations, regardless of genetic background.

Results

Patients and tumors

Eight patients (two males and six females, age 30–73 years) were included in the present study (Table 1). Final diagnoses included six PGLs, one cervical schwannoma, and one cervical neurofibroma. Pathological confirmation was obtained in two PGL and two benign nerve sheath tumors. In other PGLs, the diagnosis was based on findings

<table>
<thead>
<tr>
<th>Patient</th>
<th>Diagnosis</th>
<th>Status</th>
<th>Focality</th>
<th>Tumor volume (cm³)b</th>
<th>Gold standard</th>
<th>Spectra quality</th>
<th>Succinate</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>PGL</td>
<td>SDHD</td>
<td>Multi Vagal</td>
<td>8.2</td>
<td>Pathology</td>
<td>Good</td>
<td>Detected</td>
</tr>
<tr>
<td>2</td>
<td>PGL</td>
<td>SDHD</td>
<td>Multi Jugular</td>
<td>2.3</td>
<td>PET imaging</td>
<td>Medium</td>
<td>Detected</td>
</tr>
<tr>
<td>3</td>
<td>PGL</td>
<td>SDHB</td>
<td>Uni Carotid body</td>
<td>19.6</td>
<td>PET imaging</td>
<td>Good</td>
<td>Detected</td>
</tr>
<tr>
<td>4</td>
<td>PGL</td>
<td>SDHD</td>
<td>Multi Carotid body</td>
<td>4.4</td>
<td>Pathology</td>
<td>Uninterpretable</td>
<td>NA</td>
</tr>
<tr>
<td>5</td>
<td>PGL</td>
<td>Sporadic</td>
<td>Uni Jugular</td>
<td>2.1</td>
<td>PET imaging</td>
<td>Poor</td>
<td>Not detected</td>
</tr>
<tr>
<td>6</td>
<td>PGL</td>
<td>Sporadic</td>
<td>Uni Jugular</td>
<td>1.5</td>
<td>PET imaging</td>
<td>Poor</td>
<td>Not detected</td>
</tr>
<tr>
<td>7</td>
<td>Neurofibroma</td>
<td>Uni</td>
<td>Vagal</td>
<td>ND</td>
<td>24.9</td>
<td>Pathology</td>
<td>Good</td>
</tr>
<tr>
<td>8</td>
<td>Schwannoma</td>
<td>–</td>
<td>Uni Vagal</td>
<td>ND</td>
<td>6.7</td>
<td>Pathology</td>
<td>Medium</td>
</tr>
</tbody>
</table>

ND, not done.

a18F-FDOPA and 68Ga-DOTATATE findings in PGLs evaluated by MRS.
bTumor volumes were measured on contrast-enhanced T1-weighted MR images using OsiriX Software (v5.6, 64 bit, Geneva, Switzerland).
from images using various specific tracers. Genetic testing was performed in all but one PGL and revealed SDHD mutations in three cases (patient nos 1, 2, and 4) and SDHB in one case (patient no. 3). Tumors evaluated by MRS were localized to the jugular foramen in three cases, the retrostyloid parapharyngeal space in three cases, and the carotid body in two cases. Mean tumor volume was 8.7 ml and ranged from 1.5 to 24.9 ml.

MRS findings
Spectra quality was considered good in three cases, medium in two, poor in two, and uninterpretable in the latter case due to motion artifacts. Smaller lesions had lower spectra quality compared to larger lesions. Jugular PGL also exhibited a poorer spectra quality compared to vagal PGL and CBP. A succinate resonance peak was only detected in SDHx-related tumor patients. Succinate was detected in patient no. 3, who had a 2.3 ml jugular PGL with medium quality spectra. Two examples of 1H-MR spectra in comparison to HRMAS findings are presented in Figs 1 and 2. A spectra without succinate is shown in Supplementary Fig. 1 (see section on supplementary data given at the end of this article).

Discussion
The present study demonstrates that 1H-MRS could enable in vivo detection of succinate in SDHx-related tumors. These results emphasize that, beyond its localization
value, this imaging modality provides unique opportunities for better characterizing these tumors at a metabolomic level that is uniquely linked to their molecular signature, in these as well as other tumors. Thus, recently, it has been proposed that ex vivo detection of succinate and other metabolites could guide genetic testing (Lendvai et al. 2014, Richter et al. 2014). In recent years, it has also been demonstrated that immunohistochemistry using specific antibodies against SDH subunits is a reliable method for predicting SDH mutations (van Nederveen et al. 2009). However, these and other ex vivo techniques have some limitations because they are not applicable in cases where PGLs are treated by repeated radiation, cannot be removed surgically due to their unusual location or size, and are unable to provide continuous specific metabolite assessment that would be very useful for monitoring or predicting changes in intratumoral metabolism, tumor aggressiveness, resistant or responsive therapy, and metastatic spread. Therefore, in vivo metabolomics characterization of any tumor is becoming of paramount interest for guiding genetic, therapeutic, and outcome evaluation of cancer patients.

Metabolomics or metabolite profiling is the youngest sibling in the family of -omics fields and is growing up. Maturing right behind genomics, transcriptomics, and proteomics, metabolomics is the comprehensive analysis of small molecule metabolites (Reitman et al. 2011). Succinate is a component of the TCA cycle that serves as an electron donor to complex II. Succinate is present in the brain at ~0.5 mmol/kg (Klunk et al. 1996). Although present at such a low concentration, it contains four protons from two methylene groups that all contribute to a singlet at 2.39 p.p.m. In conventional in vivo 1D MRS experiments, this signal overlaps with resonances of glutamate and glutamine (Govindaraju et al. 2000). However, using HRMNAS, we have previously shown that SDHx-associated PGL exhibits a very low glutamate content (Imperiale et al. 2015a). Increased succinate has also been reported in human brain abscesses (Shukla-Dave et al. 2001).

Pioneering studies or hypotheses from investigations by Selak et al. (2005) showed the accumulation of succinate in SDHx-tumors. Elevated plasma succinate has even been proposed as a screening test for detecting SDHx mutation-positive individuals (Hobert et al. 2012). Detection of succinate has been also found to be very useful for classifying SDH variants of unknown etiology as pathogenic or depicting SDH deficiency without an SDHx mutation such as an SDH promoter methylation that may occur in some cases like Carney triad (Haller et al. 2014, Imperiale et al. 2015a). More recently, a significantly increased succinate:fumarate ratio has also been described in SDHx-related PGLs and proposed as a new metabolic marker of these tumors (Lendvai et al. 2014, Richter et al. 2014). Several studies have shown that succinate and possibly other metabolites, the so-called PHEO/PGL metabolomics milieu, play important and perhaps the most crucial role in the pathogenesis, behavior, and outcome of these tumors (Vicha et al. 2014). Thus, beyond SDHx mutations, disruption of the TCA cycle has been described for other mutations that predispose to PHEOs/PGLs such as isocitrate dehydrogenase type 1 (Gaal et al. 2010), fumarate hydratase (Castro-Vega et al. 2014), and the more recently described malate dehydrogenase type 2 (Cascon et al. 2015). It is also expected that the detection of other TCA enzyme mutations may play an important role in the pathogenesis of PHEO/PGL, and the use of metabolomics to uncover new PHEO/PGL-specific metabolic profiles will become crucial in novel discoveries of such mutations in the very near future.

High MRS spectra quality, demonstrated by the ability to separate resonances from important metabolites within the tumor, depends on various technical aspects such as voxel and pre-saturation band placement, pre-acquisition shimming quality, acquisition parameters, water and fat suppression, and post processing. Compared to the brain, the MRS of HNPGGs is challenging due to their anatomical location near or within bone structures and the surrounding adipose tissue, which create susceptibility artifacts and make the shimming process very difficult. Furthermore, data quality is also degraded by patient motion (head movement) and vascular pulsatility. In the present study, jugular PGL and small lesions exhibited a poorer spectra quality compared to other sites. These PGLs, which arise from the dome of the jugular vein and are located in the temporal bone, are more sensitive to susceptibility artifacts, with specific problems for optimizing shimming and fat suppression. In one CBP, motion artifacts lead to an uninterpretable spectrum. A manual shimming procedure could improve spectral quality but requires longer examinations.

In our study, smaller lesions were also found to have lower quality spectra. In these cases, the lower size of the voxel volume decreased the signal-to-noise ratio (SNR). It is possible to reduce the voxel size to 1 ml (Abdel Razek & Poptani 2013), but this requires increasing the number of excitations with a subsequent increased duration of the scan.

The present and previous studies open and strengthen a new field of in vivo metabolomics profiling in various PHEOs/PGLs. In recent years, 18F-FDG uptake has also been
shown to be strongly dependent on patient genotype. Thus, the degree of 18F-FDG uptake has also been proposed as a predictor of SDHx PHEOs/PGLs (Taieb et al. 2009, 2014b, Timmers et al. 2012, Blanchet et al. 2014). According to this lesion-based model using SUV ratio and tumor diameter, sensitivity, specificity, positive predictive value, negative predictive value, and accuracy were 80.8, 63.6, 83.1, 60.0, and 75.5% respectively (Blanchet et al. 2009). Recently, Imperiale et al. (2015) have confirmed the importance of monitoring therapeutic responses in vivo by hyperpolarized 31P-MRS (Mishkovsky et al. 2011). Hyperpolarized succinate can be produced using para-hydrogen induced polarization and was thus used as a contrast agent for MRI and MRS in preclinical studies of brain tumors (Bhattacharya et al. 2007). More recently, it has been shown that hyperpolarized (2H, 13C)-labeled glucose by dynamic nuclear polarization increases the SNR up to 10 000 times (Ardenkjaer-Larsen et al. 2003). This contrast agent could be used for the assessment of succinate and other TCA metabolites by MRS (Mishkovsky et al. 2012) and successfully applied to various hereditary PHEO/PGL that are considered as a metabolic disease.

In conclusion, in vivo metabolomics analysis may serve as an important bridge between molecular genetics and imaging. 1H-MRS could be added to the classical MR sequences for characterization of various PHEOs/PGLs, especially those related to TCA cycle impairment. Imperiale et al. (2015b) have also recently shown that 1H-MRS enables in vivo detection of catecholamines in PHEOs. The recently introduced PET/MR systems enable acquisition of MRS and PET data during a single examination, provide high-quality fusion of both modalities and potentially provide the opportunity to perform multi-voxel acquisition analysis in the setting of prospective studies.

In vivo detection of succinate shows promise in further guiding genetic testing and characterization of SDH variants, especially in the absence of available tumor samples and detection of changes in plasma succinate levels in these patients, which is currently insufficient. This approach also has the future potential of serving as an important tool for monitoring therapeutic responses while avoiding excessive radiation exposure or functional imaging techniques that are often very costly yet limited in availability. Nevertheless, the conclusions of the present study should be tested on a very large population of patients with SDHx and non-SDHx tumors.

**Supplementary data**

This is linked to the online version of the paper at http://dx.doi.org/10.1530/ERC-15-0246.

**Declaration of interest**

The authors declare that there is no conflict of interest that could be perceived as prejudicing the impartiality of the research reported.

**Funding**

This research did not receive any specific grant from any funding agency in the public, commercial or not-for-profit sector.
References


Received in final form 17 June 2015
Accepted 23 June 2015
Made available online as an Accepted Preprint 25 June 2015